Camellia  

*Camellia spp.*

**Propagation**

Camellias are usually propagated by cuttings of new growth taken just as the stem is changing from green to brown. Wound the stem, use 3000-8000 ppm IBA-talc and grow in sand:peat or peat:perlite with mist. Camellias can also be grafted or tissue cultured.

**Pests and Diseases**

The current Insect Management Guide for Commercial Foliage and Woody Ornamentals can be found at [http://edis.ifas.ufl.edu/IG012](http://edis.ifas.ufl.edu/IG012). The current Professional Disease Management Guide for Ornamental Plants can be found at [http://edis.ifas.ufl.edu/PP123](http://edis.ifas.ufl.edu/PP123).

**Algal spot** (*Cephaleuros virescens*)

**Recognition:** Silvery-grey, green or tan, raised spots or blotches with green margins on leaves are caused by algal leaf spot. Heavily infected plants may experience premature yellowing and leaf loss.  

**Contributing factors:** Abundant moisture, high temperature and direct sunlight favor infection. It seems to be most damaging on slow-growing, weakened plants.  

**Management recommendations:** Use selective pruning to increase air circulation and remove heavily infected leaves. No fungicides are recommended.

**Dieback, Twig blight or Canker** (*Colletotrichum gloeosporioides* - asexual stage, *Glomerella cingulata.*)

**Recognition:** New, succulent growth in the spring wilts and then leaves drop. Entire twigs and shoots may turn brown and die as temperatures increase. Cankers form on older wood at the base of the dead twigs. Persistent active cankers can last for
years on woody stems providing inoculum that will enter wounds and kill cuttings.

**Contributing factors:** Most severe in warm, humid conditions. On some species (e.g. *C. oleifera*), the disease will quickly kill large branches, while other species (e.g. *C. hiemalis* ‘Kanjiro’) will heal over the canker. The strain of *Glomerella* that affects camellia will not attack other plants, however, it will attack all species of camellia except *C. sinensis, C. sasanqua* and *C. oleifera* are more susceptible than *C. japonica. C. sasanqua* ‘Cleopatra’ is particularly susceptible, whereas *C. japonica* ‘Charles S. Sargent’ and ‘Governor Mouton’ are particularly resistant. A wound or natural opening is required for infection. Leaf scars are the most common point of entry. The symptoms of wilting and leaf drop may be confused with Phytophthora root rot. If root rot is present, the dieback may be even more severe.

**Management recommendations:** An integrated approach is the best way to manage this disease. Splashing water spreads this disease, so it is best to isolate diseased plants and minimize overhead irrigation and the length of time foliage and stems are wet. Organize the nursery to increase space between plants to improve air movement and sun exposure. Avoid wounding plants and take cuttings from near the tops of healthy plants. Prune out diseased twigs, shoots and cankers as soon as possible, pruning at least six inches below visibly affected areas. Remove and discard diseased plants and prunings. Disinfect pruning tools regularly during pruning and follow good sanitation practices when propagating. Chemical control is targeted in the spring when the old leaves are shedding to protect leaf scars from infection in areas that have had previous dieback. One application at this time is usually sufficient.

**Flower Blight (Ciborinia camelliae)**

**Recognition:** This disease only affects flowers in various stages of development. Irregular tan to light brown spots first appear on petals of expanding flowers, and progress to turn the entire petal, and then flower, brown with gray mycelium at the base of the petals near the calyx. Small, hard, black sclerotia develop on the mycelium and allow the fungus to survive in the soil until the next season. Flower blight can be distinguished from cold, wind, or water damage by the growth of the spots, a netted appearance with the veins darker than the surrounding tissue,
the presence of sclerotia and by the scattered appearance of the blighted flowers
(cold damage would affect flowers in a larger area).

**Contributing factors:** This disease tends to be most severe on *C. japonica* because the
flower development occurs at the same time as the release of spores. *C. sasanqua*
and other species open earlier, often before spores are released, thereby avoiding
infection. The disease is also most noticeable on light colored flowers. Abundant
moisture and temperatures around 70 F increase the growth of the organism.

**Management recommendations:** Chemicals are used only where there is a history of
severe infection, and should be targeted to the ground around and under the plants
in the fall prior to flowering. A repeat application may be required if flowering is
longer than four weeks. Keep the pathogen out of the nursery to avoid
establishment of the disease by bringing in only clean plants and flowers. New
plants from areas of infection should be bare-root to avoid the chance of sclerotia
in the soil. Prune lower branches to increase air movement and to facilitate
removal of old flowers. Collect and destroy diseased flowers.

**Wet root rot diseases (Phytophthora and Pythium spp.)**

**Recognition:** Above-ground symptoms are poor growth, thinning of the foliage, and
yellowing of leaves, with the oldest foliage affected first. These symptoms may be
one-sided on the plant. Wet rots cause a soft decay of the outer layers of roots,
which can be easily stripped off between two fingers, leaving the firm, white stele intact.

**Contributing factors:** The disease is triggered by periods of excessive soil
moisture. Conditions that favor disease development include planting too deep,
poor drainage, shallow rooting, and poor water management.

**Management recommendations:** Check roots of nursery-grown plants before planting into the landscape. Provide
adequate drainage, and reduce irrigation. Apply labeled fungicides if problem is
diagnosed early and cultural problems corrected.

**Root rots (Phytophthora, Pythium spp.; Rhizoctonia solani)**

**Recognition:** The first symptoms noticed are usually in the foliage, which turns a lighter
green, somewhat like a nutrient deficiency, then becomes yellow and wilts and
eventually falls off. Infected roots are discolored brown and rotted, with no feeder
roots, and occasionally cankers on root crown and lower stem. Plants may become
stunted and die quickly if young with a limited root system, or slowly decline if
older.

**Contributing factors:** Root rots are favored by prolonged periods of saturated soils and
moderate temperatures. Spores are moved around in water, soil, container mix and
on equipment and plant material. Infection may occur when conditions are right, but may not be noticed until hot weather when the plants have a higher water demand and the lack of roots is limiting. *C. japonica* and *C. reticulata* are the most susceptible species of Camellia to *Phytophthora*, and may be grafted on *C. sasanqua*, a more resistant species, to improve plant survival.

**Management recommendations:** Avoid overwatering, heavy mixes that retain too much water, placing pots directly on the soil and standing water. Using a porous material to cover beds, rather than a solid plastic cover, and raising the beds slightly will allow water to drain away and prevent standing water. Always use uninfested potting mix or incorporate composted bark media known to be naturally suppressive to soil pathogens. Use only new or disinfested pots. Disinfest cuttings prior to use and take only from the tops of clean stock plants. Disinfest tools and work surfaces. Do not use untreated recycled irrigation water during propagation or on susceptible cultivars. Chemical controls do not eliminate the fungus, but only prevent it from active infection. Therefore repeated applications during periods of heavy rainfall and moderate temperatures may be required.

**Aphids**

**Recognition:** Aphids have small pear-shaped bodies with paired cornicles in rear. They may or may not have wings. Aphids cluster and feed on new growth — flowers, stems and leaves. Sooty mold grows on honeydew excreted by the aphids. Several kinds and colors of aphids feed on hibiscus. (Note: photo shows cotton aphids)

**Contributing factors:** Aphids occur in the spring and throughout the growing season.

**Management recommendations:** Aphid "mummies" are a sign that natural biological control is occurring. High aphid populations may be treated with insecticidal soaps, oils, or approved insecticides.
Scale (30 species, tea scale most common)

**Recognition:** Variegation on the top of the leaf may be seen with severe infestations. The scale lives on the bottom of the leaf and in severe infestations can look fuzzy in a cluster.

**Contributing factors:** Eggs are laid when temperatures are 86° to 91°F. Depending on temperature, the tea scale life cycle takes between 45 to 65 days. An adult female will lay between 10 to 15 eggs, shrivel up, and die shortly thereafter. In warmer climates like Florida, scales reproduce continually throughout the year but in cooler climates hatching will often coincide with the warming spring temperatures.

**Management recommendations:**

**Cultural.** Pruning is provides for better coverage of chemical sprays and increased air circulation. In cooler climates small non-flowering branches growing on major limbs within the interior of the plant should be pruned between February and March.

**Chemical.** In cooler climates, spring is the best time to apply chemical insecticides as the danger of cold weather has passed and egg hatching often coincides with the warming temperatures. It is essential that thorough coverage of the leaf's under-side is attained. The addition of a sticker-spreader is an effective way to increase coverage. Repeat applications (2-3) made between seven to 10 days apart is necessary to manage a tea scale infestation. Prior to making pesticide applications, efforts should be made to insure that a current tea scale infestation is not being naturally managed by native parasites. The use of soaps and oils are preferable to insecticides because they are usually less harmful to the natural predators of tea scale. Follow the manufacturer's labeled rate for any product applied to control a pest. The current Insect Management Guide for commercial foliage and woody ornamentals can be found at [http://edis.ifas.ufl.edu/IG012](http://edis.ifas.ufl.edu/IG012).

**Biological.** Several wasps native to the United States, including *Aphytis diaspidis* and two species of *Aspidiotiphagus*, have been reported parasitizing tea scales in Florida and Georgia. The female wasp will insert a single egg into the tea scale. Parasitized scales have detectable holes chewed out in their armor by the emerging wasp and are often associated with patches of necrotic tissue.

Spider mites

**Recognition:** Tiny (less than 1/50 inch) eight-legged mites are found most commonly on the undersides of leaves. Eggs, cast skins and silken webs are also signs of mites. Mites feed with piercing-sucking mouthparts, causing the upper sides of leaves to exhibit a stippled or bronzed appearance. Common mite pests of azaleas include both southern red mites and two-spotted spider mites. Two-spotted spider mites have dark spots.
Southern red mites are named for their bright red color.

**Contributing factors:** Southern red mites are most prevalent in cool, moist, conditions, whereas two-spotted spider mites prefer hot, dry weather.

**Management:** Use a white piece of paper to monitor for mites. Check for predaceous mites as well, which are larger and move more quickly. Insecticidal soaps, horticultural oils, or approved miticides may be used to control mites when necessary.

**Nutrient Deficiencies**

**Magnesium deficiency**

**Nitrogen deficiency**

**Sources:**

6. Nutrient deficiency, spider mite damage, flower blight and algal leaf spot photos courtesy of Florida Department of Agriculture and Consumer Services, Division of Plant Industry.

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